

Specimen Guide

Guidelines

- Submit a complete clinical history including time & date of obtaining the sample
- Submit specimens in proper container or vacutainer tubes
- Each specimen must be clearly labeled as to the source & individual animal identification
- Package the specimen carefully to eliminate breakage in transit
- Samples should arrive before 3:00 PM Monday – Friday & by 9:00 am on Saturday

<p>Routine Chemistry Collect blood in sterile red top tube. Allow the blood to clot for 15-30 minutes before centrifuging at a high speed (2500 rpm) for 10 minutes. Remove the serum and transfer into a clean sterile tube. Ship overnight with an ice pack. Minimum of 1 ml of serum should be submitted for most chemistry profiles. DO NOT SUBMIT WHOLE BLOOD OR SERUM SEPARATOR TUBES FOR CHEMISTRY TESTS.</p>	<p>Hematology: Blood should be collected in EDTA tubes. Tubes MUST be at least 1/3 full. Mix samples immediately after collection. Check for clots & redraw if clots are present. Make 2 peripheral blood smears. Allow to air dry and send with the EDTA specimen. High temperatures & freezing must be avoided. Ship specimens overnight in an appropriate container.</p>
<p>Bile Acid –Liver Function Test Fasting Bile Acid – Fast patient for 8 to 12 hours, collect clot tube. Centrifuge and remove serum from clot. Check sample for hemolysis & lipemia. Redraw if necessary. Post Prandial Bile Acid – After collecting the fasting sample feed a small amount of low fat food (KD or ID is recommended). Dogs < 5 pounds feed 1 tablespoon Dogs >5 but <50 pounds feed ¼ cup Dogs >50 pounds feed ½ cup Ensure food is retained. Draw a two hour post prandial serum sample. Centrifuge and separate serum. DO NOT SUBMIT IN SERUM SEPARATOR TUBES. Random Bile Acid – Collect in red top tube. Centrifuge and submit like a routine chemistry test.</p>	<p>Coagulation: All coagulation tests must be collected in a FULL sodium citrate tube (blue top tube). The ratio of blood to anticoagulant is critical. Centrifuge specimen and remove plasma and freeze in a no-additive red top tube. Ship frozen specimen on ice packs overnight. Label specimen as citrated plasma</p>
<p>Special Chemistry Test: TSH, T4, & Progesterone - Draw samples in a sterile red top tube. Allow the sample to clot thoroughly before centrifuging. Separate serum & transfer to a clean tube. <i>Progesterone</i> testing must be shipped overnight. For Saturday delivery, please ship UPS ACTH Stimulation Test - For dogs & cats collect a baseline sample and administer 5 µg/kg IV and obtain a post sample at 1 hour later. Low Dose Dexamethasone – for canine, collect baseline sample then administer 0.01 mg/kg dexamethasone iv. Obtain post samples at 4 and 8 hours. Ionized Calcium – Draw in red top tube. Centrifuge & immediately freeze serum. Send on ice packs</p>	<p>Miscellaneous Test: Urinalysis – Fresh urine should be sent in a sterile container on ice packs. If cytology is desired, submit small amount of urine in an EDTA tube. ERD – 3 ml fresh urine sample (detection of micro-albumin levels) Coombs/Crossmatch – Send EDTA and Serum sample. Snap 4DX – 0.5 ml serum or EDTA plasma. Detection of <i>Dirofilaria immitis</i> (HW), <i>Anaplasma phagocytophilum</i> (AP) <i>Borrelia burgdorferi</i> (LY) & <i>Ehrlichia canis</i> (EC).</p> <p>Parasitology: Submit 3-5 g fresh feces in a sterile leak proof container. Please do NOT submit specimen in gloves.</p> <p>Cytology Peritoneal, thoracic, urine & other body fluids should be transported in an EDTA tube (purple top tube). Submit 2 air-dried, unstained direct smears with the sample. DO NOT send in formalin Fine needle aspirates or impression smears should be submitted as air-dried smears. DO NOT use fixative. More than one smear is preferred, with sample from 2 to 3 different areas of the lesion. Please include a complete clinical history and description of condition for all cytology submissions. DO NOT SUBMIT SYRINGES.</p>